



Certificate of Analysis

Standard Reference Material[®] 1932

Fluorescein Solution

This Standard Reference Material (SRM) is intended for use in establishing a reference scale for fluorescence intensity based upon Molecules of Equivalent Soluble Fluorophore (MESF) units [1–3]. This SRM is certified for the concentration of fluorescein with a certified purity in a borate buffer solution. The MESF scale is established for a particular set of experimental conditions by measuring the fluorescence intensity of known amounts of this SRM under a specific set of conditions (see “Instructions for Use”). A unit of SRM 1932 consists of three flamed sealed amber glass ampoules, each containing approximately 2.0 mL of fluorescein in an aqueous borate buffer.

Certified Value: The certified concentration of fluorescein given below is based on gravimetric preparation and analysis of purity by proton nuclear magnetic resonance spectroscopy that was independently verified by analysis of impurities. A summary of fluorescein purity determinations is provided in Table 2. A NIST certified value is a value for which NIST has the highest confidence in its accuracy in that all known or suspected sources of bias have been investigated or taken into account [4]. The measurands are the concentration and purity of fluorescein and the traceability is to the SI derived units for amount-of-substance concentration and mass fraction, expressed as micromoles per kilogram and percent, respectively.

Fluorescein mass fraction: 60.97 ± 0.40 ($\mu\text{mol}\cdot\text{kg}^{-1}$)
Fluorescein purity: 97.55 ± 0.64 (%)

Reference Values: Table 1 gives reference values for the molar absorption coefficient of SRM 1932 corrected for purity and fluorescence bias. NIST reference values are noncertified values which represent the best estimate of the true values based on available data; however, the values do not meet the NIST criteria for certification [4] and are provided with associated uncertainties that may reflect only measurement precision, may not include all sources of uncertainty, or may reflect a lack of sufficient statistical agreement among multiple analytical methods. The measurands are the molar absorption coefficients as listed in Table 1 as realized by the methods used. Metrological traceability is to the SI derived units for molar absorptivity, expressed as kilograms per mole centimeter.

Table 1. Reference Values for Molar Absorption Coefficient of SRM 1932 at $22.4\text{ }^\circ\text{C} \pm 0.5\text{ }^\circ\text{C}$

Wavelength (nm)	Molar Absorption Coefficient ($\text{kg}\cdot\text{cm}^{-1}\cdot\text{mol}^{-1}$)	Uncertainty ($\text{kg}\cdot\text{cm}^{-1}\cdot\text{mol}^{-1}$)
488.0	8.50×10^4	0.07×10^4
490.0	8.70×10^4	0.07×10^4
490.5	8.71×10^4	0.07×10^4
491.0	8.70×10^4	0.07×10^4

Expiration of Certification: The certification of **SRM 1932** is valid, within the measurement uncertainty specified, until **30 June 2022**, provided the SRM is handled and stored in accordance with the instructions given in this certificate (see “Instructions for Use”). The certification is nullified if the SRM is damaged, contaminated, or otherwise modified. The certification is valid only for unopened ampoules that have been stored in the dark between $2\text{ }^\circ\text{C}$ to $6\text{ }^\circ\text{C}$.

Maintenance of SRM Certification: NIST will monitor this SRM over the period of its certification. If substantive technical changes occur that affect the certification before the expiration of this certificate, NIST will notify the purchaser. Registration (see attached sheet or register online) will facilitate notification.

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Certificate Revision History on Last Page

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Overall direction and coordination of technical measurements leading to certification were performed by G.W. Kramer of the former NIST Biochemical Sciences Division.

Production and certification of this SRM were performed by P.C. DeRose and A.K. Gaigalas of the NIST Biosystems and Biomaterials Division; B. Coxon and G.W. Kramer of the former NIST Biochemical Science Division; and T.W. Vetter of the NIST Chemical Sciences Division; with assistance from D.L. Duewer, J.B. Smeller, M.B. Satterfield, D.M. Bunk, M.J. Welch, E. White V., Y. Tewari, S.A. Margolis, J.R. Sieber, L.L. Yu, and L. Wang of NIST.

Ancillary measurements were provided by G.E. Marti, F. Abbasi, and J. Weaver of the U.S. Food and Drug Administration, R.F. Vogt of the Centers for Disease Control and Prevention, and Y-z. Zhang of Molecular Probes, Inc.

Statistical consultation was provided by J. Lu of the NIST Statistical Engineering Division.

Support aspects involved in the issuance of this SRM were coordinated through the NIST Office of Reference Materials.

Source of Material⁽¹⁾: The fluorescein (2-(6-hydroxy-3-oxo-3H-xanthen-9-yl)benzoic acid, C₂₀H₁₂O₅, relative molecular mass 332.311, CAS No. 2321-07-5) powder used in SRM 1932 was prepared especially for this purpose by Molecular Probes, Inc. (Eugene, OR, U.S.; Leiden, The Netherlands), 71358, MPR, Lot W018073. Boric acid granules were obtained from Mallinckrodt (St. Louis, MO, U.S.), Lot 2549 KVTK, relative molecular mass 61.83. Sodium hydroxide pellets were from Mallinckrodt, Catalog No. 7708, Lot 7708M484721, relative molecular mass 40.00. All water used was NIST deionized water that was then subsequently passed through a second purification system (Millipore Mill-Q A10) to produce water having a resistivity ≥ 18 M Ω cm.

Determination of Fluorescein Purity: Table 2 summarizes the purity determinations of the fluorescein material used to prepare SRM 1932. The measurands are the impurities as listed in Table 2 as realized by the methods used. Metrological traceability is to the SI derived units for mass fraction, expressed as percent.

Table 2. Reference Values for Fluorescein Purity Determinations and Associated Uncertainties

Constituent	Technique	Mass Fraction (%)	Uncertainty (%)
Fluorescein	¹ H NMR ^(a)	97.55	0.64
DHBBA ^(b)	¹ H NMR ^(a) & HPLC ^(c)	1.04	0.20
Ethanol	¹ H NMR ^(a)	0.21	0.04
Methyl Isobutyl Ketone	¹ H NMR ^(a)	0.02	0.02
HOAc/Acetate	¹ H NMR ^(a)	0.02	0.01
Ethyl Acetate	¹ H NMR ^(a)	0.02	0.01
Water	Karl Fisher	0.25	0.02
Total Organics	CHO Analysis ^(d)	99.19	0.82
Potassium	FAES ^(e)	0.50	0.14
Sodium	FAES ^(e)	0.03	0.03
Chloride	Argentimetry ^(f)	0.37	0.03

^(a) 500 MHz Proton Nuclear Magnetic Resonance Spectroscopy

^(b) 2-(2',4'Dihydroxybenzoyl) benzoic acid

^(c) High Performance Liquid Chromatography and High Performance Liquid Chromatography-Mass Spectrometry, both normal and reversed phase

^(d) Elemental analyses carried out by Schwartzkopf Microanalytical Laboratory, Inc. (Woodside, NY), Atlantic Microlab, Inc. (Norcross, GA), and Galbraith Laboratories, Inc. (Knoxville, TN)

^(e) Flame Atomic Emission Spectroscopy

^(f) Halide content identified solely as chloride by X-ray Fluorescence and Inductively Coupled Plasma-Mass Spectrometry

⁽¹⁾Certain commercial equipment, instruments or materials are identified in this certificate to adequately specify the experimental procedure. Such identification does not imply recommendation or endorsement by the National Institute of Standards and Technology, nor does it imply that the materials or equipment identified are necessarily the best available for the purpose.

Information Values: Table 3 gives information values for SRM 1932. A NIST information value is a value that may be of use to the SRM user, but insufficient information is available to assess adequately the uncertainty associated with the value. Information values cannot be used to establish metrological traceability.

Table 3. Information Values for SRM 1932

pH (25 °C)	Buffer Concentration	Mass Density (22 °C)
9.48	0.10 mol·L ⁻¹	1.003 g·mL ⁻¹

Preparation of the SRM: The borate buffer solution was prepared by dissolving 18.56 g of boric acid granules in slightly less than 3 L of water. The pH was adjusted to above 9.1 using 40 mL of an approximately 3.0 mol·L⁻¹ NaOH solution made by dissolving 29.87 g of sodium hydroxide pellets in 0.250 L of water. The mixture was diluted to 3.00 L with water to give a borate buffer with a boric acid concentration of about 0.10 mol·L⁻¹.

The SRM solution was prepared in a darkened room by dissolving 0.05636 g ± 0.00004 g of fluorescein (buoyancy corrected) in 2.7134 kg ± 0.0006 kg of borate buffer solution (buoyancy corrected). The mass of fluorescein used was corrected for the fluorescein purity yielding a final fluorescein concentration of 60.97 μmol·kg⁻¹ ± 0.40 μmol·kg⁻¹. The fluorescein solution contained in an amber bottle was immediately aliquotted into ampoules that were subsequently flame-sealed. The pH and mass density of the SRM were determined from measurements on four ampoules selected randomly from the lot.

Assignment of Uncertainties: Standard uncertainty components equivalent to the estimated standard deviation were assigned for sample inhomogeneity and measurement uncertainties. These values were then combined with balance accuracy and estimated instrument method uncertainties using the root-sum-of-squares method. An expansion factor of $k = 2$ was applied so that the expanded uncertainties given in this certificate express an interval within which the true value is expected to fall with a level of confidence of approximately 95 % for a normal distribution [5].

SRM Stability: NIST monitored the stability of a prototype fluorescein solution similar to SRM 1932 for over 24 months. Within the error of the measurements, the absorbance spectrum and the fluorescence spectral radiance (as indicated by the fluorescence signal spectrum) of the prototype solution did not change. Therefore, SRM 1932, if stored in the dark between 2 °C to 6 °C, is likely to maintain its original optical properties for the duration of its certification period. NIST will validate this conclusion by periodic monitoring of the stability over the lifetime of the SRM (see “Maintenance of SRM Certification”).

INSTRUCTIONS FOR USE

CAUTION: This SRM is a solution contained in tip-sealed glass ampoules with pre-scored stems. Therefore, all appropriate safety precautions, including the use of gloves during handling, should be taken to avoid accidental breakage or spillage. Unopened ampoules should be stored in the dark between 2 °C to 6 °C in an upright position. The ampoules should **NOT** be frozen because of the possibility of breakage during freezing and thawing. Once the ampoule is opened, the solution should be used promptly with minimal exposure to any light (exposure to incandescent lighting is preferable to illumination from daylight or fluorescent lighting). Any unused solution in the ampoule should be discarded in accordance with all applicable Federal, State and Local regulations.

Opening an Ampoule: When an ampoule is opened, that area of the stem where the pre-scored band is located (around the gold band) should be wiped with a clean, damp cloth and the body of the ampoule wrapped in absorbent material. Then holding the ampoule steady and with the thumb and forefinger grasping the stem above the gold band, **minimal** thumb pressure should be applied to the stem to snap it. Correctly done, the stem should break easily where pre-scored. The use of a metal file to break the stem is **NOT** recommended.

The SRM 1932 solution should always be diluted gravimetrically at least one-hundred fold with the same buffer system used for the analyte of interest. Subsequent gravimetric dilutions can then be employed to generate a calibration curve. The calibration curve describes the relationship between fluorescence intensity, as determined by the fluorometer and the concentration of fluorescein, as determined by the gravimetric dilution. Care must be taken in making the gravimetric dilutions because uncertainties expand during this serial process. An error at one level adversely affects determinations at all subsequent levels. The calibration curve measurements should be generated starting with the lowest concentration. For best results, the conditions used for determining the calibration curve such as solution degassing, temperature, ionic strength, pH, etc., should closely match those used in the measurement of the analyte. Because fluorescence is a highly sensitive technique, great attention must be paid to the cleanliness of glassware and any other apparatus that contacts the solutions. Many plastics and gloves can contaminate samples with small amounts of highly fluorescent materials such as release agents and plasticizers. The running of blanks to check for such contamination is highly recommended.

Depending on the solution pH, aqueous fluorescein solutions are complex, rapidly equilibrating mixtures of its several forms (cation, neutral species, monoanion, and dianion). Each species has unique absorbance and fluorescence spectra. Above pH 9, aqueous fluorescein exists almost exclusively as the highly fluorescent dianion. However, as the pH of the solution is reduced, the concentration of the dianion decreases, and the concentrations of the much less fluorescent monoanion and neutral forms increase [6]. Accordingly, the sensitivity of the fluorometric assay for fluorescein also decreases, and quantitation becomes very dependent on knowing or maintaining the precise pH of the solutions during calibration as well during the assay itself. While the calibration strategy described above can work with solutions below pH 9, the uncertainties of such measurements inevitably grow larger as the solution pH is lowered.

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<p>Certificate Revision History: 05 June 2017 (Change of expiration date; editorial changes); 18 June 2012 (Change of expiration date; editorial changes); 10 December 2007 (Change of expiration date; editorial changes); 01 December 2004 (Update of absorption coefficient reference values (and related text) based on measurements corrected for bias associated with purity and fluorescence, rather than photobleaching; change of expiration date); 04 March 2003 (Original certificate date).</p>

Users of this SRM should ensure that the Certificate of Analysis in their possession is current. This can be accomplished by contacting the SRM Program: telephone (301) 975-2200; fax (301) 948-3730; e-mail srminfo@nist.gov; or via the Internet at <http://www.nist.gov/srm>.